Prevention and management of postharvest diseases with

physical means

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WG1. Prevention of food loss and food waste

The new European plant health regulation (EU 2016/2031) jointly with the



• increment by 25% of the land used for organic farming



Aspire™

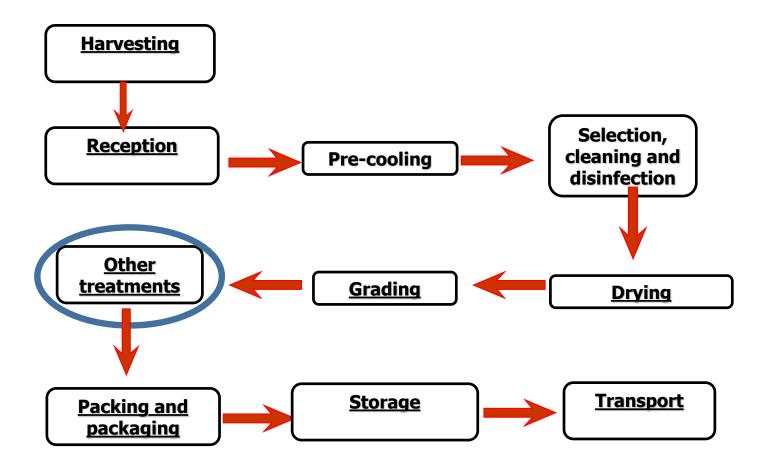


In this regard, microorganisms and Physical means (<u>Heat treatments</u>) can represent an ecological and safe strategy to adopt in agriculture

Biosave™



Fundamental Steps of Postharvest





• DA-meter

Heat treatments

- Botanicals
- Biological control agents

Postharvest control strategies

Fungicide treatment		NO!!!!
Sanitation practice		YES!!!
Alternative approach	?	YES!!!

In postharvest phases, **physical means** and in particular **heat treatments** (HTs) may represent a valid approach due to the versatility with which they can be applied





Hot Water dips, rinses or brushing

Vapour and Hot air

HTs can have a twofold effectiveness against fungal diseases: preventative and curative

Type of fruit, temperature, time of exposure to the heat source and disease to control

Heat Treatments



HOT WATER:

- immersion in HW or spraying on fruits
- T° between 45 and 60 °C for up to 10 min
- Spraying consists of a pressured spray of HW, often part of a working line where fruits are moved by brush rollers

HOT AIR:



- 12 to 96 h and 38 to 46 °C
- changing in function of the heat transfer, fruit size, and sensitivity
- Slow heat transfer (small size fruits)
- HTs if not uniform can cause fruit quality damage in firmness and <u>colour</u>

<u>CURING</u>: very simple to apply leaving bins-containing fruit for the required time under a shelter, is adopted in the global kiwi industry to control gray mold (Mari et al. 2015)

OR

2-3 days in an air atmosphere heated to temperatures higher than 30 °C and a RH higher than 90% (Palou, 2013)





HTs: Mechanisms of Action

1) Fungal pathogen inhibition

On fungal spore germination and mycelial growth on fruit surface, so reducing the microbial epiphytic population:

- Direct action
- Accumulating ROS, damage of proteins and lipids

Pathogen	T°	Time
Diversi batteri	60-70	10
Bacteri rsistenti al calore	90	30
Didymella lycopersici	50	30
F. oxysporum f.sp. dianthi	60	30
F. oxysporum f.sp.	57	30
gladioli	53	30
Pythium sp.	53	30
P. irregulare	46	20-40
P. ultimum	53	30
R. solani	50	5
Sclerotinia sclerotiorum	53	30
Verticillium albo-atrum	58	30
V.dahliae	90	30
Molti attinomiceti	100	15
Diversi virus	70-80	15
Diverse infestanti	60-70	30

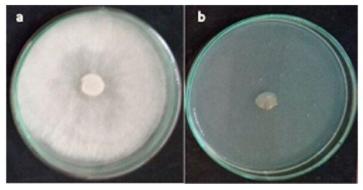
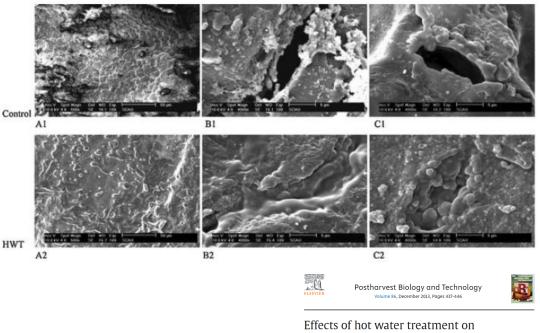


Fig. 3 The *in vitro* sensitivity of *Botrytis cinerea* to hot water treatment (HWT) using agar disk assay. a Untreated agar disk of the pathogen seeded on PDA medium and b hot water-treated (56 °C for 10 s) (Elshahawi et al., 2023)

Wet spores are more sensitive than dry ones to HTs (as well as the germinated than nongerminated ones)

2) Host structural and physiological responses

- sealing fruit entry points with epicuticolar waxes (stomata and microcracks)
- host tissues lignification (Bhuiyan et al., 2009)
- improve level of phenolic compounds and increase resistance to fungal pathogen and mycotoxin buildup (Sanzani et al., 2009)



Effects of hot water treatment on anthracnose disease in papaya fruit and its possible mechanism

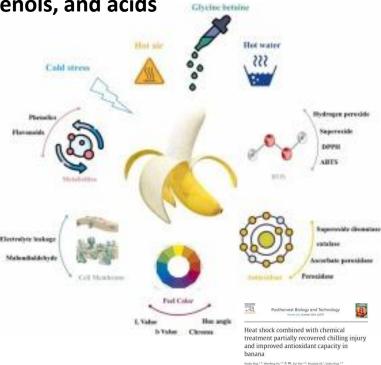
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The cracks and most stomata appeared to be partially or completely plugged by the melted wax, thereby providing a mechanical barrier against wound pathogens (54°C × 4 min)

3) Host genes regulation

- Heat Shock Proteins (HSPs) a family of proteins regulated by the heat shock transcription factors (HSTFs) that perceive abiotic stresses by activating a protection
- **Pathogenesis Related** (PR) **proteins**, significantly detectable in response to important stress factors and in particular in response to fungal pathogen infection
- Activating host antioxidant systems such as catalase (CAT), peroxidase (POD), and superoxide dismutase (SOD)
- Genes involved in fruit metabolism of sugars, polyphenols, and acids





Case studies



Postharvest losses



Brown rot

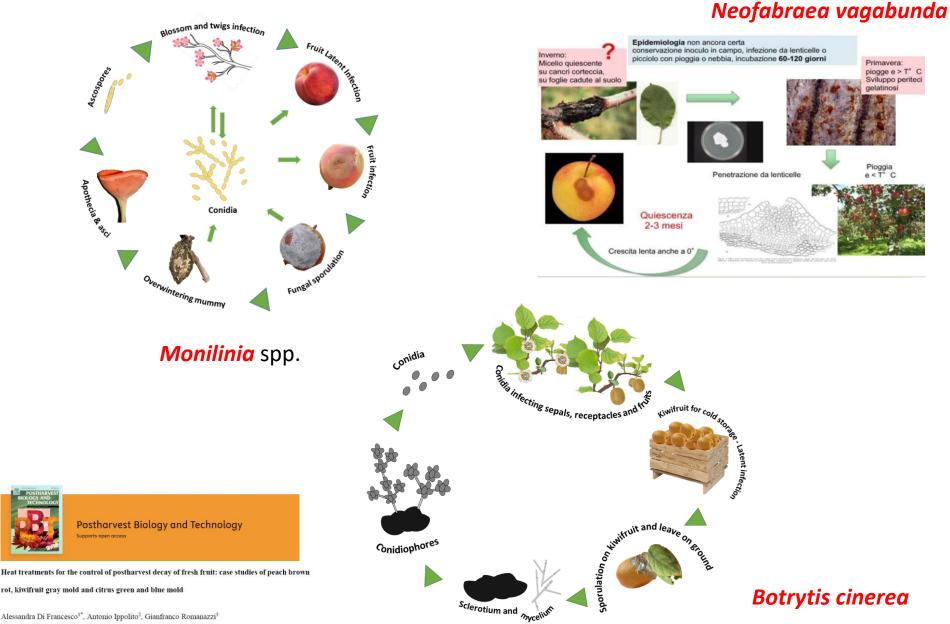
Bull's eye rot



Gray mold



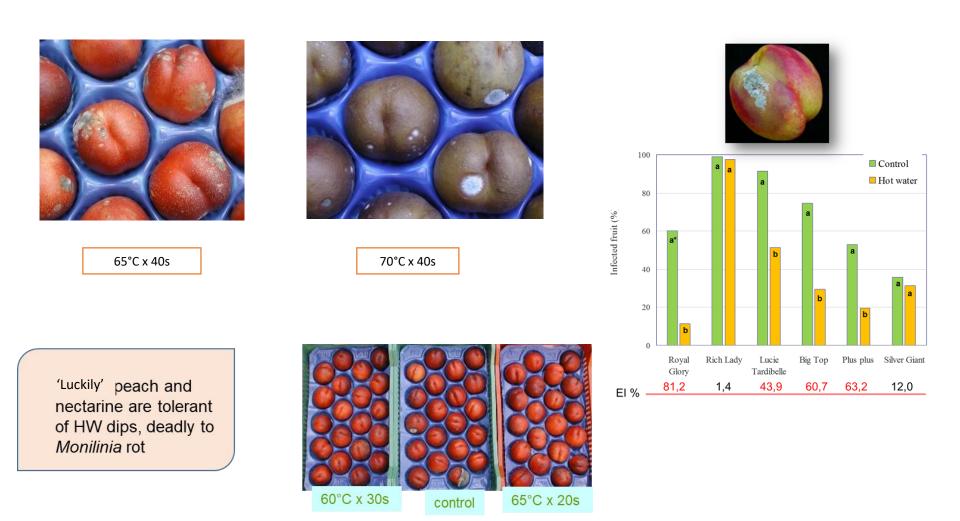
What is the main common characteristic of the three pathogens?



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LIFE+

Heat damages (skin browning) can occur when too high or too long dips are used

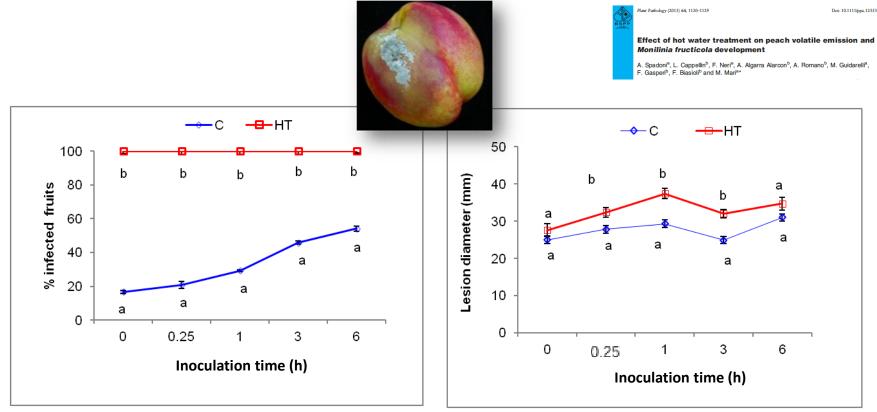




Heat treatment trials carried out by immersion in water **at 60 °C for 20 and for 60 sec** (~70% inhibition) (Spadoni et al., 2015) as curative treatment

Preventative application

Effect of hot water treatment applied **BEFORE** inoculation



HWT 60°C×60 sec Dipping inoculation (10^3 conidia/mL)

HWT 60°C×60 sec Wound inoculation (1×10^3 conidia/mL)



STIMULATORY EFFECT – after 5 days at 20°C

1° - HW treatment of peach fruit (60°C *60 sec) or tap water (control)

2° - Inoculation with *Monilinia fructicola* conidial suspension (10⁶ conidia/mL) by spray after 0 and 24h from TRT

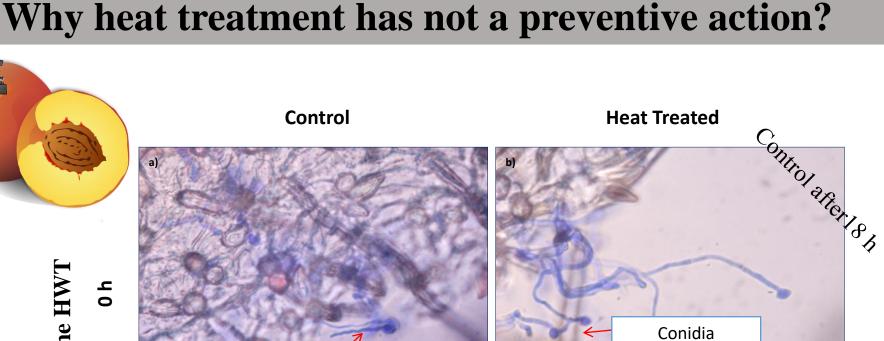
3° - HW and C peach fruit stored at 25°C for 18 h in a humid box

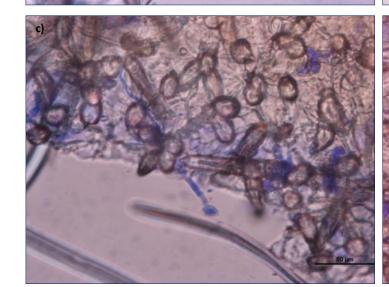
> 4°- Microscope analysis lactophenol blue (4×4 mm epidermal layer)



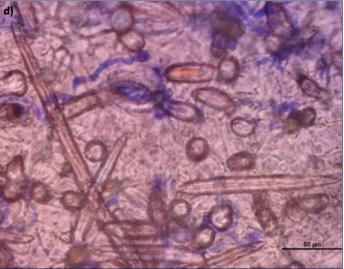
Inoculation time AFTER the HWT

24 h





Germ tube



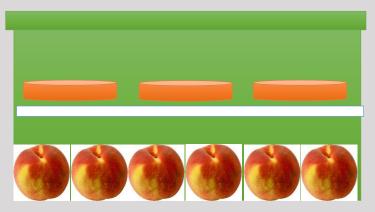
Monilinia fructicola germ tube lenght (µm) after 18 h of incubation at 25°C

Time of inoculation from the treatment (h)		Control	Heat treated		
0		58 ± 4.5 a*	$135\pm7.8~\text{b}$		
24		73 ± 6.3 a	72 ± 4.8 a		
> 2 times longer					





Inoculated Petri dishes overturned

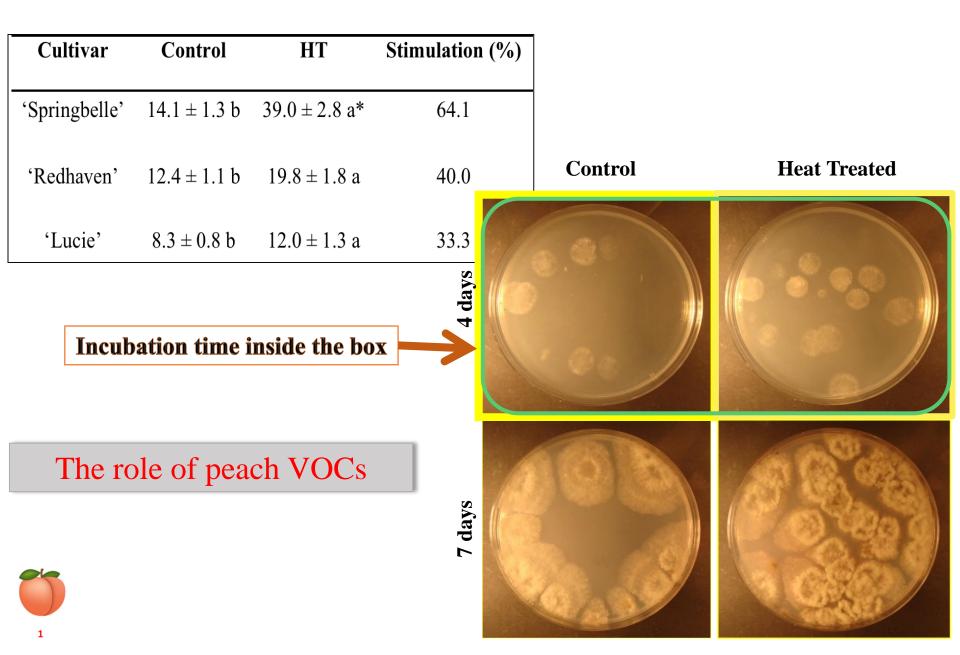


Heat treated peaches

PDA Petri dishes were spread with *M. fructicola* 10³ conidia/ mL

Cv used: Springbelle, Redhaven and Lucie Tardibelle







The role of peach VOCs

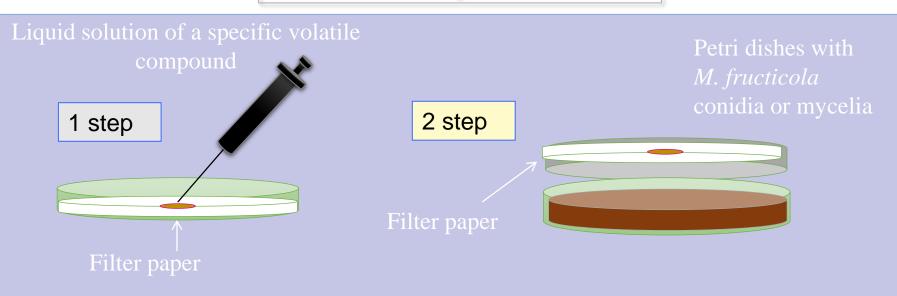
PTR-Tof-MS analysis

		CONTRO	DL	HEAT T	REATED		
Ion sum formula	Annotation	MEAN	SD	MEAN	SD	P-value	P<0.05
		(ppbv)	(ppbv)	(ppbv)	(ppbv)	1-value	1 <0.05
CH5O+	Methanol	95.7	76.5	84.4	54.6	0.755	
C2H5O+	Acetaldehyde	27.5	22.4	445.5	294.7	0.003	*
C2H7O+	Ethanol	5.1	3.5	143.0	96.3	0.003	*
C2H5O2+	Acetate fragment	6.3	1.1	16.5	9.8	0.018	*
C10H17+	Monoterpenes	3.6	2.0	4.4	1.7	0.887	



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The role of peach VOCs



	Control	Acetaldehyde	Ethanol	
	Control	0.6 ppm	0.2 ppm	
CFUs	48.2 ± 6.2 a	$77.3\pm2.4~\mathbf{b*}$	57.5 ± 6.4 a	
Ø colony (mm)	33.6 ± 1.1 a	32.0 ± 3.2 a	$39.0\pm0.4~\textbf{b}$	



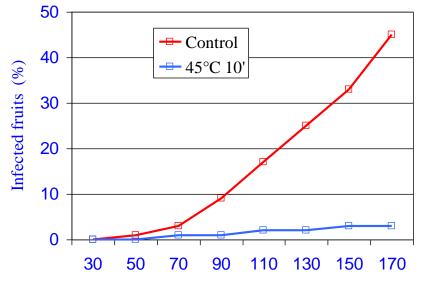
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80 Control HW 60 Infected fruit (%) а а 40 b а b 20 0 F1 F2 F3 F4 After 150 days at 0°C 47,7% 57,4% 62,3% 51,4% EI %

Hot water treatment









45°C × 10 min

Hot Water Enzyme Effect in Apple

Scientia Horticulturae Volume 227, 3 January 2018, Pages 181-186



Research Paper

B.

Defense response against postharvest pathogens in hot water treated apples

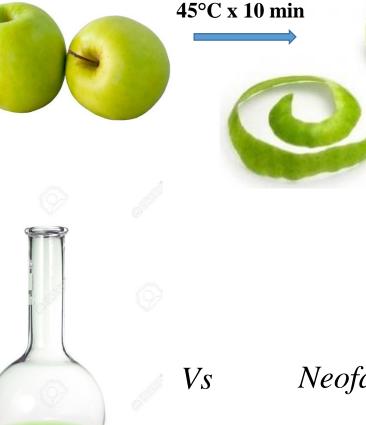
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0 - 3 - 6 – 24 h from HWT

Neofabraea vagabunda pathogenic enzymes







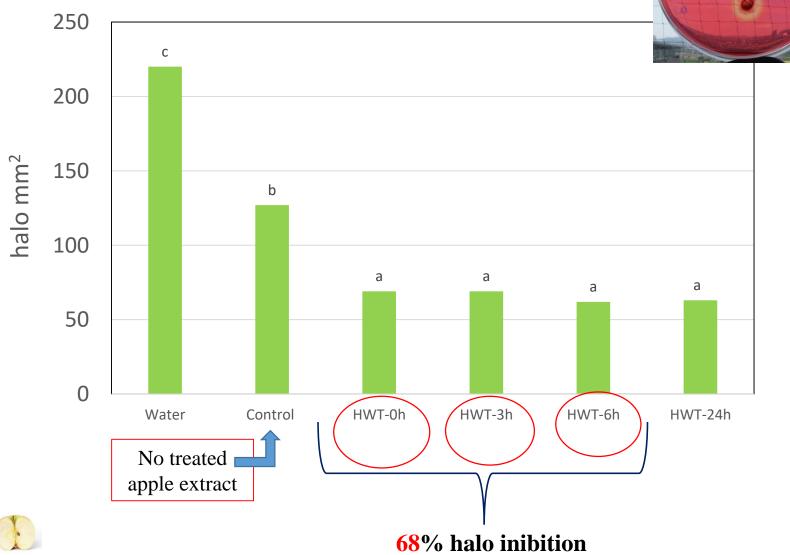


Research Paper

Defense response against postharvest pathogens in hot water treated apples

Alessandra Di Francesco º, Marta Mari º, Roberta Roberti b 🞗 🖾

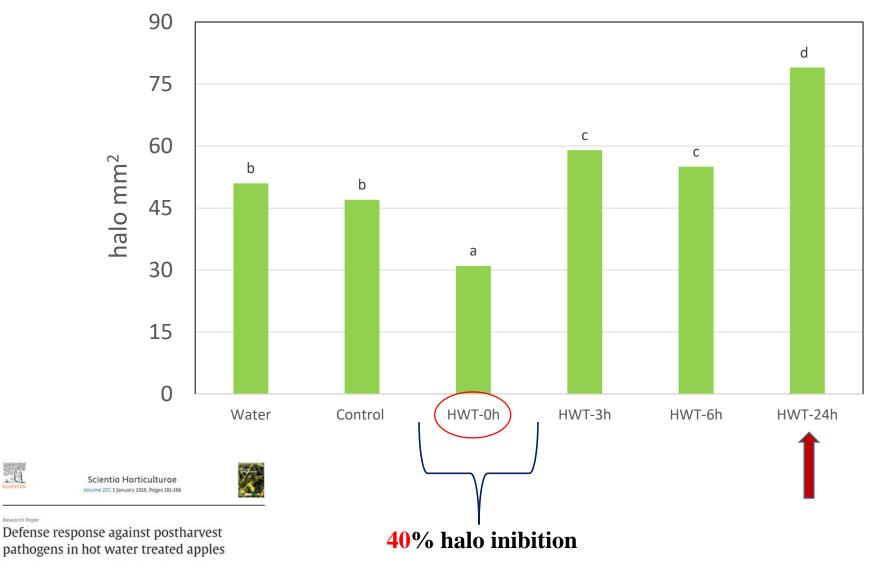
Endo-1,4-β-glucanase activity (cellulase)





2

Polygalacturonase activity



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Research Paper



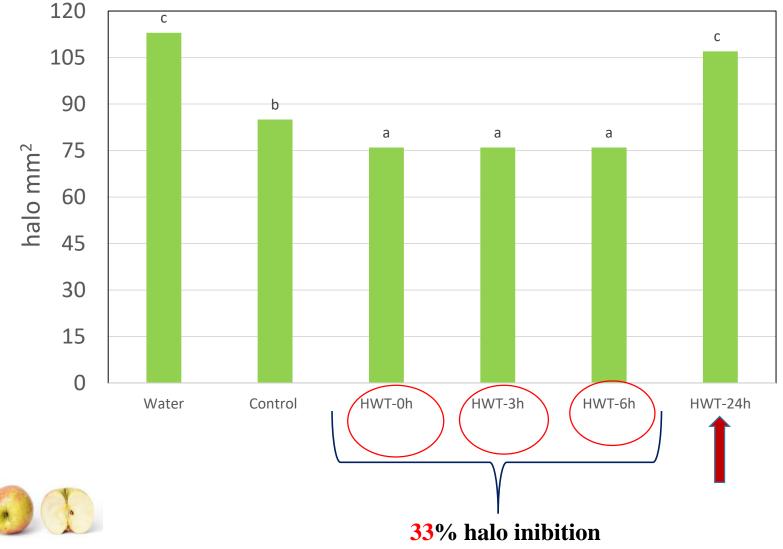


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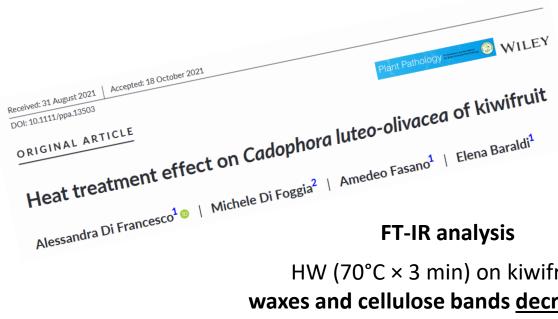
Polymethyl-galacturonase activity (pectinase)







completely inhibited gray mold at the stem end of kiwifruit (Koukounaras et al., 2008)





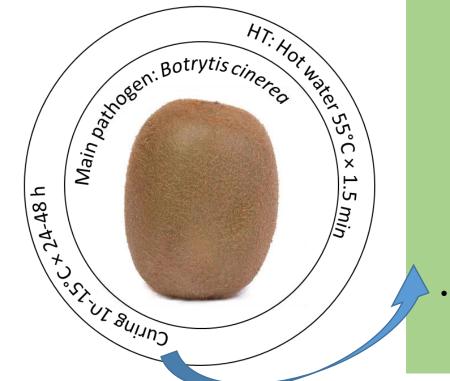
HW (70°C × 3 min) on kiwifruit skin: waxes and cellulose bands <u>decreased</u>, while phenolics, flavonoids, and glucose bands <u>increased</u>



Heat treatments for the control of postharvest decay of fresh fruit: case studies of peach brown

rot, kiwifruit gray mold and citrus green and blue mold

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3

Beneficial effect of curing:

• chitinase, phenylalanine ammonia-lyase,

polyphenoloxidase

• phenolic compounds and suberin in stem plugs and

pericarp (Ippolito et al., 1995; Wurms, 2005).

longer curing treatments (72-96 h) did not induce any

further significant reduction of infection



Tate

- Safe (no fruit residue no waste, water disposal)
 - Non selective vs fungal pathogens



• Easy to apply

• Economic (recovery of heat for cooling/storage

systems)



- The development of **time × temperature combination** remains the main challenge to keep fruit quality and resistance during postharvest phases
- In some cases, HTs do not have an improving effect on fruits (preventative)

However, the Research must always be kept active to try to find the best combinations of alternative protection systems, given the great emergence of new fungal pathogens

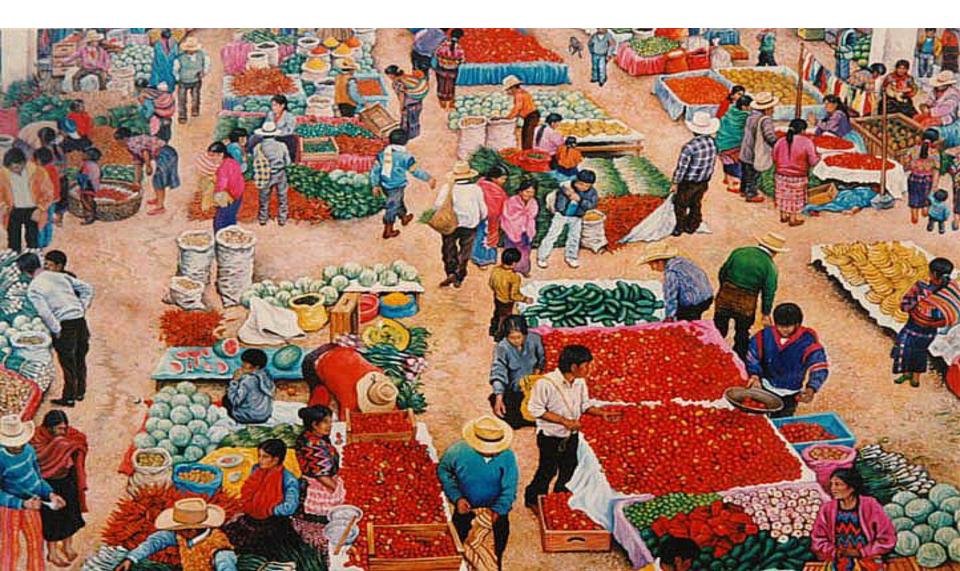
Aiming to ensure

Environmental sustainability

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Thanks for your attention





What is your opinion now on the strategies for preventing pre- and postharvest losses?